



GC-MS Analysis and In-silico Activity Evaluation of Isolated Compounds from *Nigella sativa* Seeds on Topoisomerase Alpha II

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Abstract

Plant-based medicines and naturally isolated compounds receive higher demands for the treatment of various ailments including cancer. Fewer side effects and higher therapeutic potential give more scope for drug discovery from herbals. *Nigella Sativa* or Kalonji seeds are well-known medicinal plants in the traditional system of medicine. The essential oil derived from *N. sativa* has notable biological characteristics. Even so, the essential oil of *N. sativa* contains several insoluble elements whose activities remain largely unexplored. The compounds identified using GS-MS/MS analysis of the extract identified the presence of around 32 phytoconstituents and literature shows that the isolated compound has potential activity for treating cancer. The *in-silico* docking results confirm the significant topoisomerase alpha II inhibitory activity of these compounds, especially propionic acid, 2-[(2-hydroxy-5-nitrobenzylidene) amino] -3-(1H-indol-3-yl)-, methyl ester with docking score of -10.4. Further isolation and *in-vitro*, *in-vivo* activity evaluation is needed to confirm the anticancer activity.

Keywords:

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1. Introduction

Nigella sativa L., belonging to the Ranunculaceae family, is a renowned botanical species with significant therapeutic properties. It is extensively used in several traditional medical systems, such as Unani, Siddha, and Ayurveda. *N. sativa* is used for the treatment of many chronic ailments, including diabetes, asthma, hypertension, cardiovascular disease, and cancer [1], [2], [3]. Previous investigation on *N. sativa* (NS) has identified and examined the phytochemical components of the plant, as well as explored their pharmacological effects both in laboratory settings and in living organisms. The NS seed has many bioactive phytochemical compounds, like thymoquinone, thymohydroquinone, carvacrol, isoquinolines ρ -cymene, 4-terpineol, t-anethole, nigellicine, nigellidine, and longifolene [4]. The contents of the NS seed show exceptional therapeutic characteristics, which include antiparasitic, antibacterial, antipyretic, anti-inflammatory, analgesic, antioxidant, and anticancer effects. In addition, several [5] research have shown that the extract from NS seeds may be used to inhibit cough, slow down the development of cancer, break down kidney stones, and treat polio, diarrhea, stomach discomfort, and flatulence [6]. The active components found in NS seeds have a significant impact on preventing the development of cancer and causing the death of cancer cells in different types of cancer, such as cervical cancer, breast cancer, fibrosarcoma, prostate cancer,

colon cancer, hepatic cancer, pancreatic cancer, blood cancer, renal cancer, skin cancer, and lung cancer [7]. As an example, dimethylbenz[a]anthracene, which is a key ingredient in NS extract, inhibited the development of skin cancer in mice. Treatment with NS extract resulted in a delay in the creation of papillomas [8]. NS seeds contain specific fatty acids that demonstrated cytotoxicity in Ehrlich ascites carcinoma, sarcoma-180 cells, and Dalton's lymphoma ascites [9].

The combination of *N. sativa* extract and an oxidative stress agent demonstrated significant anticancer effects in MCF-7 breast cancer cells [10]. Studies reported that administering NS volatile oil orally hinders the development of colon cancer and restrains the growth of colon cancer cells in rats during the post-initiation stage. NS extract inhibited toxicity, renal oxidative stress, and carcinogenesis produced by KBrO₃ [11]. Thymoquinone, an active element in NS, triggered apoptosis in myeloblastic leukemia HL-60 cells by caspase-8 activation, independent of the p53 protein [12]. NS seeds have a distinctly acrid flavor and are used as dietary supplements in candy and several other comestible items. In addition, NS seeds are ingested with honey, and milk and included in baked goods or pastries. *Nigella sativa* (NS) seeds are rich in vital nutrients including essential fatty acids, antioxidants, and a variety of vitamins. They also include important minerals such as calcium, iron, potassium, selenium, magnesium, and zinc [13], [14]. They

also contain a variety of fatty acids, including unsaturated ones like linoleic and oleic acid, as well as saturated ones like palmitic acid, palmitoleic acid, myristic acid, linolenic acid, myristoleic acid, margaroleic acid, margaric acid, stearic acid, eicosenoic acid, behenic acid, arachidic acid, and lignoceric acid [15]. Furthermore, NS seeds contain sterols like as β -sitosterol, stigmasterol, campesterol, Δ^7 -avenasterol, and lanosterol [16]. The kalonji seed has a chemical composition that is well-suited for the efficient treatment of several ailments. Nevertheless, these seeds also comprise vital oils and very lipophilic substances, and these insoluble elements have not been well examined, either in laboratory experiments or in living organisms.

With around 2.3 million current cases identified in 2020, breast cancer stands as a prevalent form of cancer among women globally. Breast cancer originates in the epithelial lining of the mammary glands and ranks as the second most prevalent form of cancer globally [17]. The treatment of breast cancer involves a range of approaches, such as surgical intervention, chemotherapy, and radiation. However, these techniques are associated with high costs, significant discomfort, and potential adverse consequences [18]. Although there have been advancements in early detection methods and systemic treatment options, the majority of breast tumors exhibit medication resistance [19]. Therefore, it is essential to create innovative and economical treatment modalities or medications that have minimum adverse reactions. Topoisomerase α II (Topo α II) is a well-established anticancer target. The activity of this protein activity has been linked to the sensitivity of the cancer cells, such as increased Topo α II expression is the indication of more aggressive breast cancer and that leads to disease-related deaths.

The well-known anticancer medication, doxorubicin has the Topo α II inhibitory activity [20]. The diverse chemical structures of the phytochemicals present in the novel formulation are expected to have synergistic activity and therefore prevent and treat breast cancer. In this study, we have analyzed the *in-silico* topoisomerase alpha II inhibitory activity of the phytochemicals identified using GC-MS analysis of the novel *Nigella Sativa* formulation using the software Discovery Studio.

2. Materials and Methods

2.1. Gas Chromatography-Mass Spectroscopy Analysis

The GC-MS/MS study was performed by an Agilent 8890 gas chromatography system with a mass selective detector connected to the front injector. For the chromatography, an HP- 5 MS Ultra Inert capillary column (30 m 0.25 m, film thickness M) was employed. The temperature of the oven was programmed to begin at 60 °C, rise to 310 °C at a rate of 10 °C/min, and then be maintained at 200 °C for 5 minutes. The temperature for the injection was fixed at 280°C. Helium was employed as the carrier gas with a split ratio of 30:1, a sample injection volume of 1 μ l, an injector temperature of 250 °C, and an ion source temperature of 280 °C. The compound's composition percent was calculated using the GC peak areas. Substances were analysed by GC-MS using an Agilent GC 8890 gas chromatography and an Agilent MS/MS 7000 D triple quadrupole mass spectrometer. The identical GC conditions as those listed for GC analysis were applied, along with a 1000 amu column. The unknown component's spectrum was

compared to the mass spectra and relative retention periods of the known components found in the 17th version library data of the National Institute of Standard and Technology (NISTGC-MS/MS) system. The GC-MS ran for around 32 minutes in total.

2.2. Molecular Docking

From the data bank of protein, topoisomerase alpha II (RCSB PDB-1CM8) was downloaded, and Discovery Studio 4.1 software was used for docking.

2.3. Preparation of the protein

To find potential issues, protein is produced in this stage. Missing loops are automatically built and fixed, and the side chains of missing residues are optimized.

2.4. Docking by C-Docker Modes

An interface to the LibDock program created by Diller and Merz is the Dock Ligands (LibDock) protocol. A carbon atom is preferred as the acceptor or donor atom for the H bond in polar hot spot compounds. The docking process begins with the calculation of the receptor Hot Spot file. By using docking simulation, the 31 compounds in formulation obtained from GC-MS analysis were created and geometrically optimized. The compounds under investigation were docked using imatinib as a reference into the crystal structure of a topoisomerase alpha II homodimer attached to DNA. The chemicals that were observed were stored in mol2 format. The molecule was imported into Discovery Studio 4.1 after the hydrogen bonds were added. The obtained PDB format was stored. For each position, the CDOCKER interaction energy was computed [21].

3. Results & Discussion

3.1. G-C MS Analysis

Figure (1) depicts the NSF's GC-MS profile, which includes 32 peaks for biomolecules. The phytochemicals, their retention time, peak area percentage, and molecular weight are shown in Table 1. Our findings match previous literature that demonstrated these phytoconstituents have anti-inflammatory, antioxidant, and anticancer properties. 2,3-dihydro-3,5-dihydroxy-6-methyl-4H-Pyran-4-one (DDMP) is showing antioxidant properties [22], and octanoic acid, a linear saturated fatty acid is exhibiting anticancer activity against colorectal, mammary gland and skin cancer [23]. Dodecanoic acid moiety has anticancer activity in conjugate forms and was reported to have the characteristics of apoptosis induction and cell cycle arrest [24]. Glucobrassicin is a major glucosinolate with anticancer effects and is a precursor of indole-3 carbinol [25].

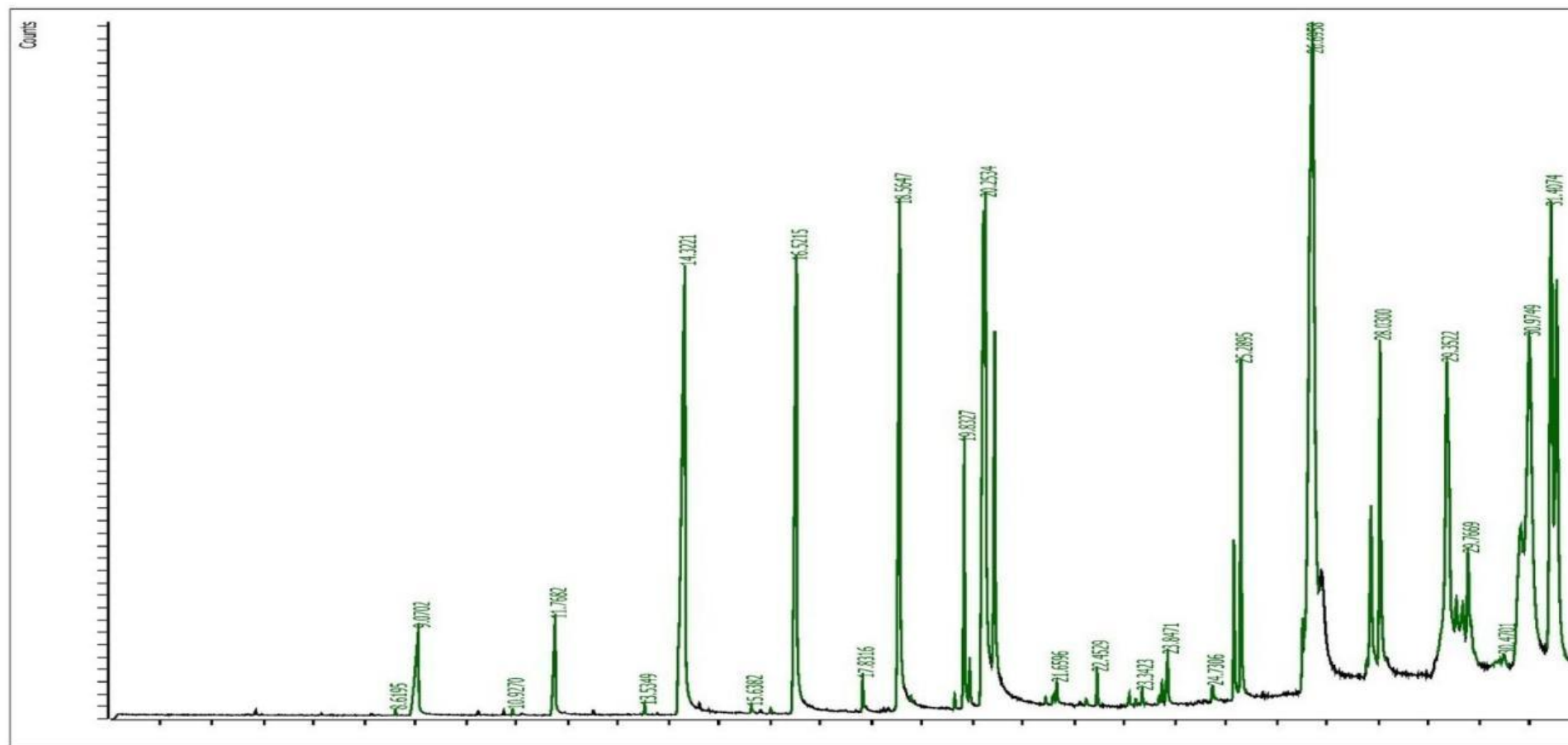


Figure 1

Figure 1. The GC-MS chromatogram of formulation showing compounds with high peak height and peak width.

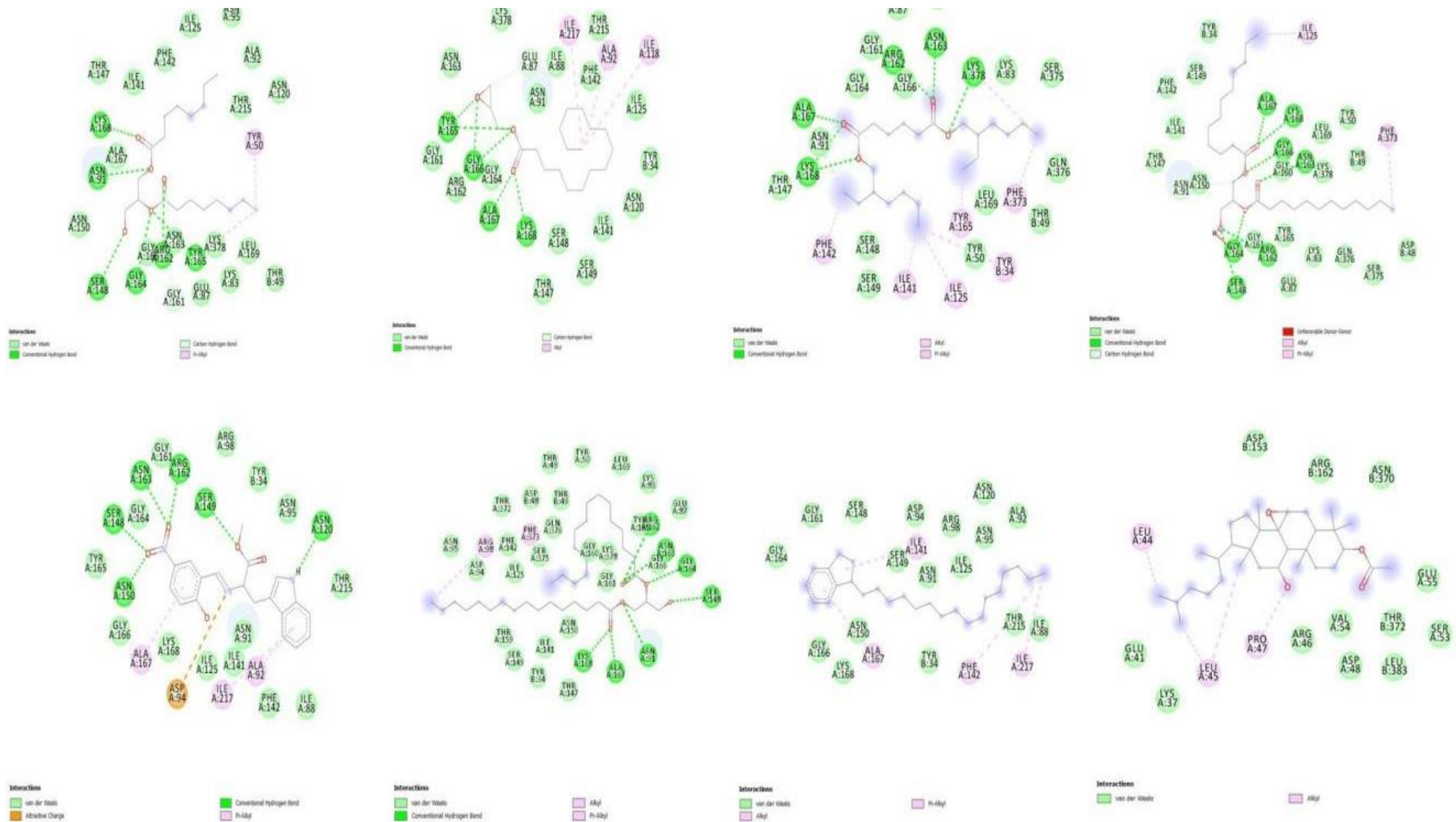
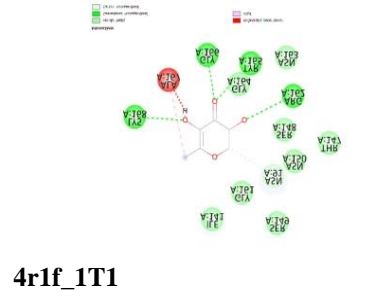
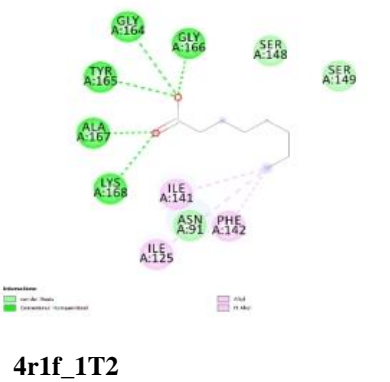
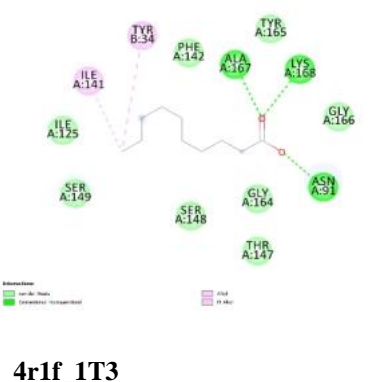
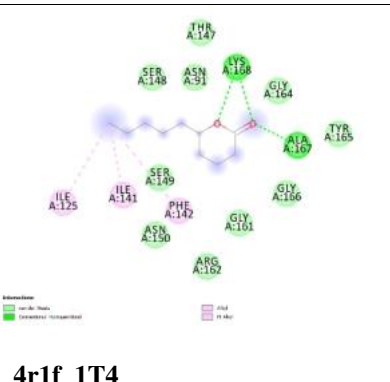


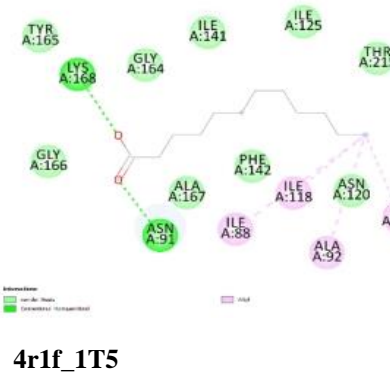
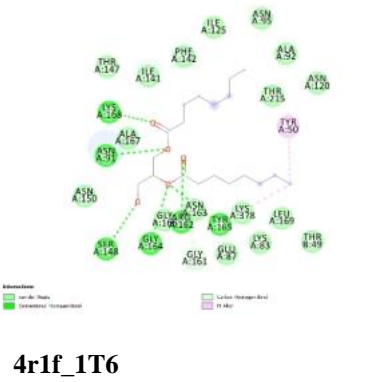
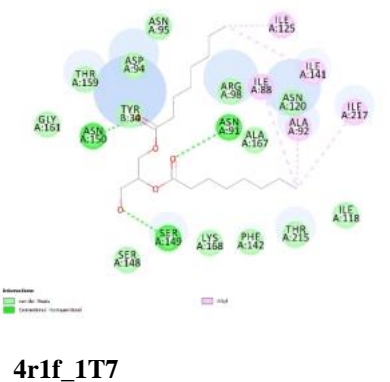
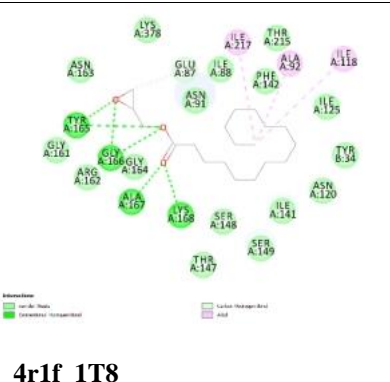
Figure 2: 2 D Binding interaction images of compounds binding interaction > -7 with Topoisomerase alpha II after docking using Discovery studio LibDock program

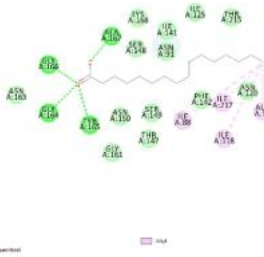
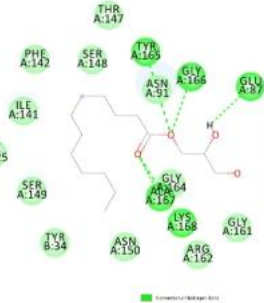
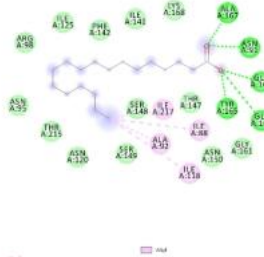
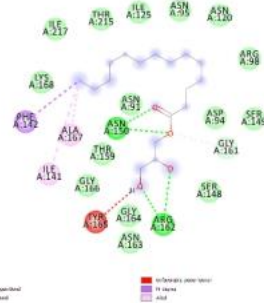
Table 1. List of phytochemicals identified in *Nigella Sativa* Virgin coconut oil formulation, their retention time and peak area% with Structural Interaction with Topoisomerase Alpha II

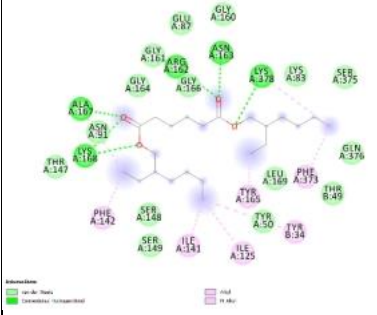
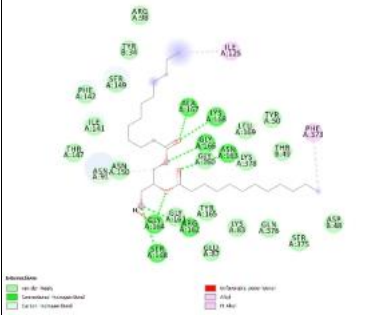
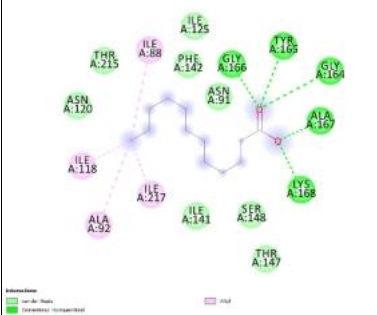
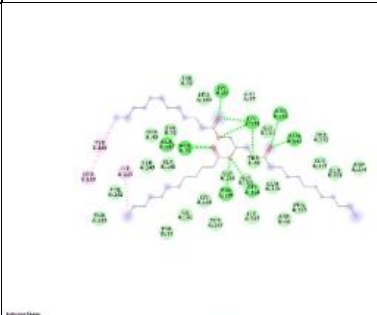
| Ligand | Lib Dock Score | Binding Interaction |
|-----------|----------------|---------------------|
| 4r1f_1T1 | 123.21 | -6.5 |
| 4r1f_1T2 | 131.41 | -5.2 |
| 4r1f_1T3 | 126.74 | -5.6 |
| 4r1f_1T4 | 126.67 | -6.1 |
| 4r1f_1T5 | 125.99 | -6.2 |
| 4r1f_1T6 | 125.55 | -7.5 |
| 4r1f_1T7 | 122.67 | -6 |
| 4r1f_1T8 | 122.57 | -7.3 |
| 4r1f_1T9 | 119.99 | -6.5 |
| 4r1f_1T10 | 119.51 | -6.3 |
| 4r1f_1T11 | 118.35 | -6.5 |
| 4r1f_1T12 | 118.14 | -6.2 |
| 4r1f_1T13 | 116.58 | -7.5 |
| 4r1f_1T14 | 112.76 | -7.6 |
| 4r1f_1T15 | 111.03 | -6.2 |
| 4r1f_1T16 | 110.59 | -6.4 |
| 4r1f_1T17 | 109.67 | -7.7 |
| 4r1f_1T18 | 108.37 | -4.4 |
| 4r1f_1T19 | 106.93 | -3.9 |
| 4r1f_1T20 | 104.99 | -5.9 |
| 4r1f_3T1 | 91.29 | -6.9 |
| 4r1f_3T2 | 90.99 | -6.5 |
| 4r1f_3T3 | 90.83 | -10.4 |
| 4r1f_3T4 | 85.99 | -6.6 |
| 4r1f_3T5 | 52.6 | -4.1 |
| 4r1f_3T6 | 85.89 | -7 |
| 4r1f_3T8 | 84.63 | -7.6 |
| 4r1f_3T7 | 85.77 | -10.4 |
| 4r1f_3T9 | 84.56 | -7.2 |
| 4r1f_3T10 | 96.52 | -4.7 |
| 4r1f_3T11 | 76.66 | -6.9 |
| 4r1f_3T12 | 145.23 | -7 |
| 4r1f_3T13 | 92.81 | -6.9 |

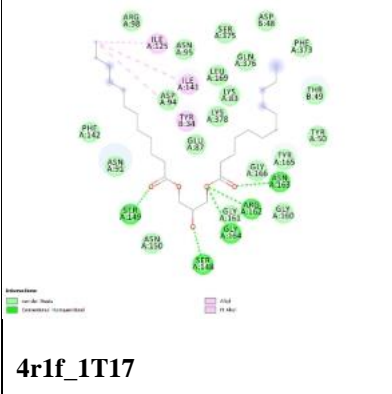
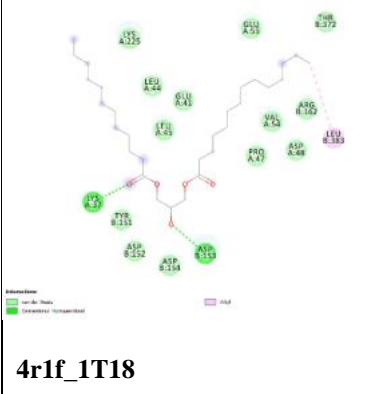
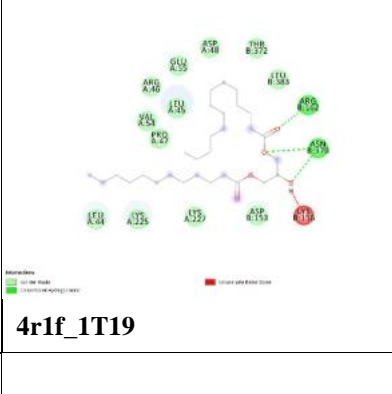
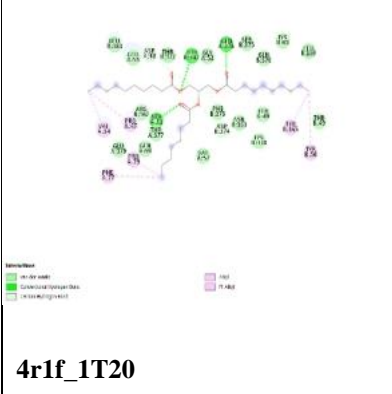

Table 2. List of phytochemicals identified in *Nigella Sativa* Virgin coconut oil formulation, their retention time and peak area% with Structural Interaction with Topoisomerase Alpha II

| Sl No | Component RT | Compound Name | Structural Interaction with Topoisomerase Alpha II | CAS# | Formula | Component Area | Match Factor | Area % Max. Area |
|-------|--------------|---|---|------------|--|----------------|--------------|------------------|
| 1 | 8.6194 | 4H-Pyran-4-one, 2,3-dihydro-3,5-dihydroxy-6-methyl- |  <p>4r1f_1T1</p> | 28564-83-2 | C ₆ H ₈ O ₄ | 4113529.3 | 69.0 | 0.2299 |
| 2 | 9.0941 | Octanoic acid |  <p>4r1f_1T2</p> | 124-07-2 | C ₈ H ₁₆ O ₂ | 225463747.7 | 91.7 | 12.6 |
| 3 | 11.7562 | n-Decanoic acid |  <p>4r1f_1T3</p> | 334-48-5 | C ₁₀ H ₂₀ O ₂ | 81520938.0 | 90.6 | 4.555 |
| 4 | 13.5349 | 2H-Pyran-2-one, tetrahydro-6-pentyl- |  <p>4r1f_1T4</p> | 705-86-2 | C ₁₀ H ₁₈ O ₂ | 7377379.8 | 75.8 | 0.4122 |

| | | | | | | | | |
|---|---------|---------------------------|---|-----------|--|-------------|------|--------|
| 5 | 14.2920 | Dodecanoic acid |  <p>4r1f_1T5</p> | 143-07-7 | C ₁₂ H ₂₄ O ₂ | 505136806.5 | 94.8 | 28.23 |
| 6 | 15.6321 | Glycerin, 1,2-dicaprylate |  <p>4r1f_1T6</p> | 502-54-5 | C ₁₁ H ₂₂ O ₄ | 5878657.7 | 71.3 | 0.3285 |
| 7 | 16.4733 | Tetradecanoic acid |  <p>4r1f_1T7</p> | 544-63-8 | C ₁₄ H ₂₈ O ₂ | 122396309.8 | 93.3 | 6.839 |
| 8 | 17.8314 | Glycidyl palmitate |  <p>4r1f_1T8</p> | 7501-44-2 | C ₁₉ H ₃₆ O ₃ | 23744070.8 | 78.5 | 1.327 |

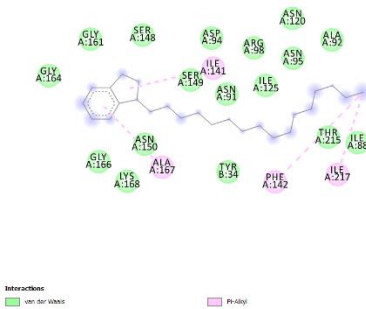
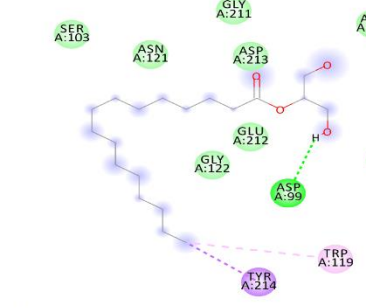
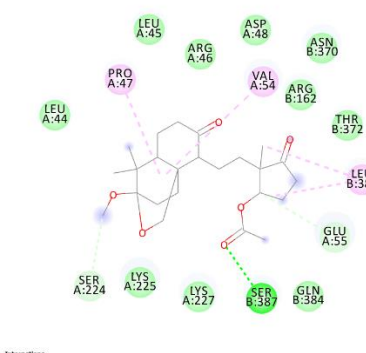
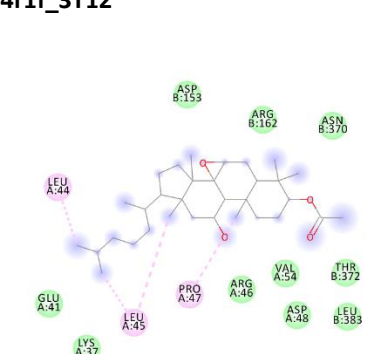
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|----|---------|---|--|----------|----------|--------------|------|--------|
| 9 | 18.5345 | n-Hexadecanoic acid |  <p>4r1f_1T9</p> | 57-10-3 | C16H32O2 | 18564854.4.2 | 93.1 | 10.37 |
| 10 | 19.8265 | Dodecanoic acid, 2,3-dihydroxypropyl ester |  <p>4r1f_1T10</p> | 142-18-7 | C15H30O4 | 97270568.2 | 92.0 | 5.435 |
| 11 | 20.4034 | Octadecanoic acid |  <p>4r1f_1T11</p> | 57-11-4 | C18H36O2 | 94390816.2 | 92.8 | 5.275 |
| 12 | 21.6534 | Tetradecanoic acid, 2,3-dihydroxypropyl ester |  <p>4r1f_1T12</p> | 589-68-4 | C17H34O4 | 11411251.7 | 74.6 | 0.6377 |

| | | | | | | | | |
|--------|---------|---|--|----------------|--------------|------------------|------|-------|
| 1 3 | 22.4526 | Hexanedioic acid, bis(2-ethylhexyl) ester |  <p>4r1f_1T13</p> | 103- 23-1 | C22H42O 4 | 26668553. 8 | 87.4 | 1.49 |
| 1 4 | 23.8469 | Dodecanoic acid, 1-(hydroxymethyl)- 1,2-ethanediyl ester |  <p>4r1f_1T14</p> | 17598- 94-6 | C27H52O 5 | 43280746. 9 | 67.4 | 2.419 |
| 1 5 | 25.2892 | 1,3-Dioctanoin |  <p>4r1f_1T15</p> | 1429- 66-9 | C19H36O 5 | 22180873 0.1 | 73.5 | 12.39 |
| 1 6 | 26.6955 | Dodecanoic acid, 1,2,3-propanetriyl ester |  <p>4r1f_1T16</p> | 538- 24-9 | C39H74O 6 | 17895521 01.6 | 68.5 | 100 |

| | | | | | | | | |
|----|---------|--|--|------------|--|--------------|------|--------|
| 17 | 28.0297 | Rac-glycerol-1,3-dilaurate |  <p>4r1f_1T17</p> | 539-93-5 | C ₂₇ H ₅₂ O ₅ | 209688486.1 | 89.3 | 11.72 |
| 18 | 29.7666 | 1-Dodecanoyl-3-myristoylglycerol |  <p>4r1f_1T18</p> | 91925-73-4 | C ₂₉ H ₅₆ O ₅ | 34655715.5 | 79.0 | 1.937 |
| 19 | 30.9686 | 1-Dodecanoyl-3-myristoylglycerol |  <p>4r1f_1T19</p> | 91925-73-4 | C ₂₉ H ₅₆ O ₅ | 1033093471.4 | 65.3 | 57.73 |
| 20 | 31.4074 | 2-(Octanoyloxy)propane-1,3-diyl bis(decanoate) |  <p>4r1f_1T20</p> | 33368-86-4 | C ₃₁ H ₅₈ O ₆ | 640212656.5 | 78.8 | 35.78 |
| 21 | 10.9270 | Phenol, 2-methyl-5-(1-methylethyl)- |  <p>4r1f_3T1</p> | 499-75-2 | C ₁₀ H ₁₄ O | 3975996.0 | 75.8 | 0.1834 |

| | | | | | | | | | |
|--------|---------|--|-----------------|--------------|------------|------------|------|--------|--|
| | | | <p>4r1f_3T1</p> | | | | | | |
| 2 2 | 18.7810 | Ethanol, 2-(9,12-octadecadienyloxy)-, (Z,Z)- | <p>4r1f_3T2</p> | 17367-08-7 | C20H38O2 | 5149133.1 | 61.0 | 0.2375 | |
| 2 3 | 19.6464 | Propionic acid, 2-[(2-hydroxy-5-nitrobenzylidene)amino]-3-(1H-indol-3-yl)-, methyl ester | <p>4r1f_3T3</p> | 1000296-35-4 | C19H17N3O5 | 8897456.6 | 60.1 | 0.4103 | |
| 2 4 | 19.9409 | 7-Methyl-Z-tetradecen-1-ol acetate | <p>4r1f_3T4</p> | 1000130-99-6 | C17H32O2 | 27898099.4 | 72.4 | 1.287 | |

| | | | | | | | | |
|--------|---------|--|-----------------|------------------|----------------|-----------------|------|------------|
| 2 5 | 20.2113 | 9,12-Octadecadienoic acid (Z,Z)- | <p>4r1f_3T5</p> | 60-33-3 | C18H32O2 | 59431563 8.2 | 93.1 | 27.41 |
| 2 6 | 20.2534 | 9-Octadecenoic acid, (E)- | <p>4r1f_3T6</p> | 112-79-8 | C18H34O2 | 51187695 0.9 | 78.1 | 23.61 |
| 2 7 | 21.4372 | Propionic acid, 2-[(2-hydroxy-5-nitrobenzylidene)amino]-3-(1H-indol-3-yl)-, methyl ester | <p>4r1f_3T7</p> | 10002 96-35-4 | C19H17N3 O5 | 5180063. 1 | 55.0 | 0.238 9 |
| 2 8 | 21.6055 | Hexadecanoic acid, 1-(hydroxymethyl)-1,2-ethanediyl ester | <p>4r1f_3T8</p> | 761-35-3 | C35H68O5 | 11459065 .0 | 70.8 | 0.528 4 |

| | | | | | | | | |
|----|---------|---|--|--------------|----------|------------|------|--------|
| 29 | 23.2101 | 1H-Indene, hexadecyl-2,3-dihydro- |  <p>4r1f_3T9</p> | 55334-29-7 | C25H42 | 4862930.5 | 53.5 | 0.2243 |
| 30 | 23.3423 | Hexadecanoic acid, 2-hydroxy-1-(hydroxymethyl)ethyl ester |  <p>4r1f_3T10</p> | 23470-00-0 | C19H38O4 | 11648572.7 | 74.7 | 0.5372 |
| 31 | 24.7306 | 8,14-Seco-3,19-epoxyandrostane-8,14-dione, 17-acetoxy-3.beta.-methoxy-4,4-dimethyl- |  <p>4r1f_3T11</p> | 1000195-87-0 | C24H36O6 | 21912324.8 | 74.0 | 1.01 |
| 32 | 30.4701 | 7,8-Epoxylanostan-11-ol, 3-acetoxy- |  <p>4r1f_3T12</p> | 1000187-60-9 | C32H54O4 | 38696002.5 | 65.5 | 1.784 |

Bio-assay-guided fractionation of Hexadecenoic acid and its activity in colorectal cancer cells showed significant MUHASINA et al., 2023

cytotoxicity in HCT-116 cell lines [26]. The essential oil component of NS, Phenol, 2-methyl-5-(1-methylethyl)-

commonly known as Carvacrol also reported to have anticancer properties [27]. Table 1 also lists the chemical makeup of the active ingredients and their interactions with topoisomerase alpha II.

3.2. Molecular docking

Based on their predicted behavior, we selected 32 phytoconstituents from GC-MS data for docking investigations using the Discovery Studio Lib Dock program. All these compounds interacted with the active site of the targeted protein topoisomerase alpha II. Although there are 10 molecules (Figure 2) having significant interactions with protein that range from -7 to -10.4 and significant roles score for LibDock that range from 84.56 to 145.23. The topoisomerase alpha II receptor served as a molecular docking target for the screening of plant phytochemicals. Figure 2 displays the top ten active substances along with their docking scores, and 2D molecular docking interactions with the topoisomerase alpha II protein.

4. Conclusions

The evidence that we acquired on NS Formulation indicates that leveraging knowledge of conventional medicinal systems may facilitate the bioprospecting, identification, development, and commercialization of novel medicinal sources that are both safe and efficient. Within this context, we have created NS formulations that exhibit varying phytoconstituents, determined by GC-MS/MS analysis of oil and the current study provides evidence that these natural compounds from NS formulation have significant topoisomerase alpha II inhibitory activity. Further investigations into NSF have the potential to provide more evidence for its efficacy in targeted therapeutic applications for cancer.

Conflict of interests

Authors confirm that there is no conflict of interest regarding this study.

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