



## Vitamin D as a marker of oocyte quality and a marker of clinical pregnancy rate after ICSI

*Enas Mostafa Mohammed, Dina Hisham Hussain, Emam Abd Allah, Ahmed Mahran, MH Ibrahim, Ayman Abdelmeged*

*Faculty of Medicine, Minia University, Minia, Egypt.*

### Abstract

Vitamin D plays an important role in Human Reproduction. This work aimed to evaluate the role of Vitamin D as a marker of oocyte quality and a marker of clinical pregnancy rate after ICSI. The study population included 88 women who went into their cycles of intra-cytoplasmic sperm injection (ICSI) treatment. The demographic parameters of the patients were evaluated. Follicular and serum 25(OH) D concentrations were measured. The following parameters were taken into consideration: The ability of the oocyte to form a good quality embryo after ICSI, Positive hCG rate after embryo transfer (ET), measured 14 days after embryo transfer. , Clinical pregnancy rate after ET is defined as the presence of an intrauterine sac with an embryonic pole demonstrating cardiac activity at 7 weeks of gestation and continuing pregnancy up to 12 weeks of gestation. The FF 25(OH)D ranged from 11.1 to 25.1 (ng/ml) with a mean FF 25(OH)D  $\pm$  SD of 18.7 $\pm$ 4.6. The Serum 25(OH) D ranged from 20.1-29.8 with a mean Serum 25(OH)D  $\pm$  SD of 25.7 $\pm$ 3.7. The positive pregnancy test was found in 32 patients representing a 36.4% success rate. Good Quality embryos were obtained on the second day of the culture in 40 patients which represent 45.5%. Ongoing pregnancy at 12 weeks was obtained in 28 patients which represents 31.8%. There was a highly statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding weight and BMI, p-value < 0.001. There was a statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding Age and height, p-value =0.002 and 0.007 respectively. There was a highly statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding the Day 3 serum FSH, Number of retrieved Oocytes, Number of fertilized Oocytes, Number of transferred embryos, Number of frozen Embryos and fertilization rate, p-value <0.001. There was a statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding the Day of embryo transfer (day five transfer was the best, p-value = 0.014). There was a highly statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding the FF 25(OH) D and Serum 25(OH) D, P value < 0.001. There was a highly statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding the Embryo quality and ongoing pregnancy at 12 weeks, P value < 0.001. There was a highly statistically significant difference between those who had a good Embryo quality and those who did not have good Embryo quality regarding the level of FF 25(OH) D and Serum 25(OH) D, p-value < 0.001. There was a highly statistically significant difference between those who had an ongoing pregnancy at 12 weeks and those who did not have an ongoing pregnancy at 12 weeks regarding the level of FF 25(OH) D and Serum 25(OH) D, p-value < 0.001. Follicular Fluid 25(OH) D and Serum 25(OH) D levels are markers of oocyte quality and a marker of clinical pregnancy rate after ICSI.

**Keywords:** vitamin D, follicular fluid and serum vitamin D, ICSI outcome.

**Full length article** \*Corresponding Author, e-mail: [hhimam@yahoo.com](mailto:hhimam@yahoo.com); [anjaz3036@gmail.com](mailto:anjaz3036@gmail.com)

### 1. Introduction

The well-known actions of vitamin D include calcium and phosphorus homeostasis in addition to the support of bone Mineralization, while its deficiency leads to an increased risk of muscle weakness, osteoporosis, and bone fractures [1]. Recent years have witnessed a shift in focus to the non-skeletal benefits of vitamin D; in this latter

context, an accruing body of literature attests to the relevance of vitamin D to reproductive physiology [2]. Vitamin D receptors (VDR) have been identified in the female reproductive tissues including human ovarian, endometrial, and fallopian tube epithelial cells, placenta, and decidual cells [3]. Vitamin D via its receptor can modulate ovarian steroidogenesis [4]. Additionally, data from clinical

reports suggest that lower vitamin D level is associated with gestational diabetes and preeclampsia. Expression of VDR is also increased during pregnancy [4, 5]. Vitamin D can contribute to the restoration of the menstrual cycle and endometrial proliferation [7, 8] promote the development of follicles [9], ameliorate primary dysmenorrhea [10], and reduce the occurrence of uterine fibroids [11]. All these mentioned effects of vitamin D led some authors to consider vitamin D as a steroid hormone [12]. However, the relationship of vitamin D levels with ICSI outcomes remains controversial. Although Polyzos et al., 2014 reported that women with deficient serum vitamin D had a lower clinical pregnancy rate compared to women with normal levels of vitamin D [13], other studies led to opposite conclusions [14-19]. Because of these conflicting reports, and given that vitamin D deficiency is not correlated with ovarian stimulation characteristics or with markers of embryo quality [16,20]. It has been postulated that vitamin D deficiency may negatively affect pregnancy rates in IVF with an effect mediated through the endometrium [13, 16, 17]. In both the epithelial and stromal cells of the endometrium, vitamin D induces and up-regulates the transcription of HOXA 10, a gene essential for implantation with increased expression during the window of implantation [20, 21]. Conversely, when a transfer of synchronous euploid blastocyst is performed, the vitamin D status does not impact the outcomes [15]. Most of the published studies on vitamin D and assisted reproduction technologies (ART) have reported outcomes related to clinical pregnancy rates or live birth rates. However, these studies were not designed to evaluate the direct relationship of local-intra-follicular vitamin D levels with the ability of the individual oocyte to undergo fertilization and subsequent embryo development [14, 15, 16, 19, 23, 24]. This work aimed to evaluate a possible relationship between the vitamin D level in follicular fluid and blood, and the fertilization rate (FR), embryo quality, chemical pregnancy, clinical pregnancy, and ongoing pregnancy up to 12 weeks gestation after ICSI.

## 2. Patients and Methods

### 2.1 Patients' selection

The study protocol was approved by the Ethics Committee of the Department of Obstetrics and Gynecology, Faculty of Medicine, and Minia University. This is a prospective observational study that was conducted in the Department of Obstetrics and Gynecology, Faculty of Medicine, Minia University during the period from July 2019 to December 2020. The study recruited patients referred for assisted reproduction treatment (ART) cycles to two of Minia's private centers for assisted reproduction under the supervision of the principal investigator according to study protocol and written informed consent was obtained from each patient after explanation of the procedure. The study population included 88 women who went into their cycles of intra-cytoplasmic sperm injection (ICSI) treatment. All included patients fulfilled the following inclusion criteria: their ages between 21 and 38 years, Their body mass indexes (BMI between 18 and 29 kg/m<sup>2</sup>), day 3 serum FSH levels <12 IU/L, their menstrual cycles are regular between 24 and 35 days, no previous history of ovarian surgery and all are well unexplained infertility. While exclusion criteria included those with moderate to severe

male factor infertility, Poor responders. None of the patients received hormonal therapy in the previous 3 months and none of them received vitamin D supplementation before controlled ovarian hyper-stimulation (COH).

### 2.2 Induction Protocol and follow up

All patients received standard controlled ovarian hyperstimulation (COH) with recombinant FSH (r-FSH) under pituitary suppression with a GnRH agonist. Oocyte retrieval was performed using trans-vaginal ultrasound 34 to 36 h after hCG injection. Intracytoplasmic sperm injection (ICSI) was performed using standard procedures and the embryos were transferred 2, 3, or 5 days later. The luteal phase was supported with 40 mg progesterone administration by daily injection. A pregnancy test was carried out on Day 14 after embryo transfer. Two weeks later, a transvaginal ultrasound was performed to confirm pregnancy. Study endpoints: Fertilization rate (FR) was calculated as the number of fertilized oocytes relative to the number of retrieved oocytes. Good quality embryos were defined as those at the 4 to 6-cell stage on Day 2. The good-quality embryos meeting these criteria were either transferred (ET) to the recipients or frozen. Biochemical pregnancy was defined by the presence of  $\beta$ -HCG >50 mIU/mL without ultrasound evidence of a gestational sac. Clinical pregnancy was defined by the presence of a gestational sac. The clinical pregnancy rate was defined by the ratio of the clinical pregnancy cases relative to the embryo transfer cases.

### 2.3 25(OH) D Measurement

25-Hydroxyvitamin D is the primary circulating form of vitamin D and remains stable throughout the menstrual cycle [25]. Due to its stability, serum 25(OH)D concentration is viewed as the best indicator of vitamin D status [26]. Heparinized serum samples and follicular fluid samples from all patients were obtained on the day of oocyte retrieval and kept frozen at -80 °C until the measurement was performed. 25(OH)D level was measured using the chemiluminescent immunoassay (CLIALIAISON®) 25 OH Vitamin D TOTAL Assay (REF 310600) DiaSorin Inc., Stillwater, MN, USA. Vitamin D deficiency was defined by the Institute of Medicine (IOM) and the Endocrine Society Clinical Practice Guidelines [27, 28] as serum 25(OH) D level < 20 ng/ml. The follicular fluid vitamin D norm remains unknown.

### 2.4 Follicular fluid (FF) collection

FF samples were collected according to the strict procedure described by Ciepiela P et al., 2015 [29]. To collect clear follicular fluid and to avoid multiple vaginal punctures and the numerous flushing of the needle with culture medium after every follicular puncture, as well as to minimize the risk of vaginal bleeding, we decided to include in the study FF only from the first nearest (available) aspirated (lead) follicle of each ovary. Consequently, vitamin D levels were representative for the lead follicle only, and not for the entire cohort of follicles from any given patient. Each ovarian follicle was aspirated independently and collected in a separate test tube to identify the matched single cumulus-oocyte-complex (COC). This approach was chosen to avoid cross-contamination from the flush medium or the FF of other

follicles. Test tubes with more than one COC were excluded from the study. In each case collected, FF was checked afterward for red blood cells. Fluids with red blood cells were disqualified. FF samples with matched mature metaphase II (MII) oocytes were centrifuged at  $10,000\times g$  for 10 min, and the supernatants were reallocated and stored at  $-80^{\circ}$ .

### 2.5 Intracytoplasmic sperm injection (ICSI)

This study included only women undergoing ICSI to allow a precise cumulus–oocyte–complex evaluation, especially about its maturity/degradation before fertilization. Furthermore, during conventional IVF insemination, there are significant events that take place, such as sperm penetration through layers of supporting granulosa cells, sperm membrane breaching, and fusion; thus, IVF requires competent oocytes and capable spermatozoa. All these events are circumvented by the ICSI procedure. While ICSI may affect the integrity of an oocyte, the literature supports the concept that oocyte ICSI degeneration is operator-independent [30].

#### 2.5.1 Embryo assessment

Embryos underwent regular embryo assessment as described by the 2011 Istanbul consensus [30]. A top-quality embryo (TQE) on the second day of the culture was defined as an embryo with four symmetrical, non-fragmented blastomeres [31].

### 2.6 Outcome measures

The primary aim of this study was to determine whether local intra-follicular vitamin D correlates with ICSI/ET results. In particular, to examine development competence of oocytes, the following parameters were taken into consideration:

- 1- Ability of the oocyte to form a good quality embryo after ICSI
- 2- Positive hCG rate after embryo transfer (ET), measured 14 days after embryo transfer.
- 3- Clinical pregnancy rate after ET is defined as the presence of an intrauterine sac with an embryonic pole demonstrating cardiac activity at 7 weeks of gestation.
- 4- Continuing pregnancy up to 12 weeks of gestation.

### 2.7 Statistical method

The analysis of the data was carried out using the IBM SPSS version 25 statistical package software. The normality of the data was tested using the Kolmogorov-Smirnov test and Shapiro-Wilk test. Data were expressed as mean  $\pm$  SD and minimum and maximum range for parametric quantitative data and by median (IQR) for non-parametric quantitative data, in addition to both number and percentage for qualitative data. Analyses were done between the two groups for parametric quantitative data using the Independent Samples T-test and non-parametric quantitative data using the Mann-Whitney test. In contrast, the Chi-square test was used to compare categorical variables. Correlation between variables was done using Pearson's correlation for continuous variables and Spearman's correlation for ordinal variables. Receiver Operating Characteristic (ROC) curve was used to determine the AUC, sensitivity, specificity, PPV, NPV, and accuracy of the Vit. D-level pregnancy and embryo quality.

Mohammed et al., 2023

A P-value less than 0.05 was considered statistically significant.

## 3. Results

### 3.1 Patients' demographic data

Eighty-eight patients were included in the study. Their age ranged from 21 to 38 with a mean age  $\pm$  SD of  $28.5\pm 5.3$ . Their weight ranged from 45 to 90.9 KG with a mean weight  $\pm$  SD of  $63.5\pm 12.5$ . Their height ranged from 1.5-1.8 M with a mean height  $\pm$  SD of  $1.6\pm 0.1$ . Their Body Mass Index (BMI) ranged from 18 to 29.2 with a mean BMI  $\pm$  SD of  $23.6\pm 3.4$ . The interquartile range of their parity was 0 to 2, with a mean of 1. Twenty-four of the patients were having primary infertility and sixty-four were having secondary infertility. The interquartile range of their duration of infertility was 2 to 4, with a mean of 3 (Table 1).

### 3.2 Intra-cytoplasmic Sperm Injection Data

Day 3 Serum FSH (IU/L) ranged from 5.1 to 11.6 with a mean level  $\pm$  SD of  $9.2\pm 2.2$ . Forty (45.5%) of the embryos were transferred on day two, 32(36.4%) of the embryos were transferred on day three, and 16 (18.2%) of the embryos were transferred on day five. The interquartile range of the Number of retrieved Oocytes was 4 to 5, with a mean of 4. The interquartile range of the Number of fertilized Oocytes was 2 to 5, with a mean of 3. The fertilization rate ranged from 50 to 100 with a mean level  $\pm$  SD of  $70.7\pm 14.8$ . The interquartile range of the Number of transferred Embryos was 1 to 1, with a mean of 1. The interquartile range of the Number of Frozen Embryos was 1 to 4, with a mean of 1 (Table 2). Thirty-two (36.4%) have positive pregnancy tests on day 14 ( $\beta$ -HCG  $>50$  mIU/mL), and 56 (63.6%) have negative pregnancy tests on day 14 ( $\beta$ -HCG  $>50$  mIU/mL) (Table 2).

### 3.3 Serum 25(OH) D and Follicular fluid level

The FF 25(OH)D ranged from 11.1 to 25.1 (ng/ml) with a mean FF 25(OH)D  $\pm$  SD of  $18.7\pm 4.6$ . The Serum 25(OH) D ranged from 20.1-29.8 with a mean Serum 25(OH)D  $\pm$  SD of  $25.7\pm 3.7$ . (Table 3).

### 3.4 Pregnancy outcome

The positive pregnancy test was found in 32 patients representing a 36.4% success rate. Good Quality embryos were obtained on the second day of the culture in 40 patients which represent 45.5%. Ongoing pregnancy at 12 weeks was obtained in 28 patients which represents 31.8% (Table 4).

### 3.5 Vitamin D and other clinical and laboratory data

There was a strong positive correlation between FF 25(OH)D and serum 25(OH)D ( $r = 0.958$ ,  $p < 0.001$ ). There was a positive or strong positive correlation between FF 25(OH)D and Age ( $r = 0.214$ ,  $p = 0.046$ ), weight ( $r = 0.373$ ,  $p < 0.001$ ), BMI ( $r = 0.396$ ,  $p < 0.001$ ), Day 3 Serum FSH ( $r = 0.627$ ,  $p < 0.001$ ), and Number of transferred Embryos ( $r = 0.345$ ,  $p < 0.001$ ). While, there was a negative or strong negative correlation between FF 25(OH)D and Parity ( $r = -0.284$ ,  $p = 0.007$ ), Number of Retrieved Oocytes ( $r = -0.381$ ,  $p < 0.001$ ), Number of Fertilized Oocytes ( $r = -0.592$ ,  $p < 0.001$ ), Number of Frozen Embryos ( $r = 0.623$ ,  $p < 0.001$ ), BHCG on day 14 ( $r = -0.633$ ,  $p < 0.001$ ), Fertilization rate ( $r = -0.432$ ,  $p < 0.001$ ), Clinical Pregnancy ( $r = -0.525$ ,  $s$

<0.001), Embryo Quality ( $r = -0.471$ ,  $s < 0.001$ ), and Ongoing Pregnancy at 12 weeks ( $r = 0.412$ ,  $s < 0.001$ ).

There was a positive or strong positive correlation between Serum 25(OH)D and weight ( $r = 0.434$ ,  $p < 0.001$ ), height ( $r = 0.290$ ,  $p = 0.006$ ), MBI ( $r = 0.417$ ,  $p < 0.001$ ), Day 3 Serum FSH ( $r = 0.626$ ,  $p < 0.001$ ), Number of transferred Embryos ( $r = 0.315$ ,  $p = 0.003$ ). While, there was a negative or strong negative correlation between Serum 25(OH)D and parity ( $r = -0.226$ ,  $p = 0.034$ ), the Number of Retrieved Oocytes ( $r = -0.654$ ,  $p < 0.001$ ), Number of Fertilized Oocytes ( $r = -0.664$ ,  $p < 0.001$ ), Number of Frozen Embryos ( $r = -0.679$ ,  $p < 0.001$ ). BHCG on day 14 ( $r = -0.656$ ,  $p < 0.001$ ), Fertilization rate ( $r = -0.498$ ,  $p < 0.001$ ), Clinical Pregnancy, ( $r = -0.525$ ,  $s < 0.001$ ), Embryo Quality ( $r = -0.450$ ,  $s < 0.001$ ), Ongoing Pregnancy at 12 weeks ( $r = -0.435$ ,  $s < 0.001$ ) (Table 5).

### 3.6 Demographic data and Clinical pregnancy rate

There was a highly statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding the weight and BMI,  $p$  value  $< 0.001$ . There was a statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding the Age and height,  $p$  value  $= 0.002$  and  $0.007$  respectively (table 6).

### 3.7 Intra-cytoplasmic Sperm Injection (ICSI) data and Clinical pregnancy rate

There was a highly statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding the Day 3 serum FSH, Number of retrieved Oocytes, Number of fertilized Oocytes, Number of transferred embryos, Number of frozen Embryos and fertilization rate,  $p$  value  $< 0.001$ . There was a statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding the Day of embryo transfer (day five transfer was the best,  $p$  value  $= 0.014$ ) (table 7).

### 3.8 Vitamin D and Clinical pregnancy rate

There was a highly statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding the FF 25(OH)D and Serum 25(OH)D,  $P$  value  $< 0.001$  (table 8). Embryo quality, ongoing pregnancy, and Clinical pregnancy rate. There was a highly statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding the Embryo quality and ongoing pregnancy at 12 weeks,  $P$  value  $< 0.001$  (table 9 and Figure 2).

#### 3.8.1 Vitamin D and embryo quality

There was a highly statistically significant difference between those who had a good Embryo quality and those who did not have good Embryo quality regarding the level of FF 25(OH)D and Serum 25(OH)D,  $p$  value  $< 0.001$  (table 10 and figure 3).

#### 3.8.2 Vitamin D and ongoing pregnancy at 12 weeks

There was a highly statistically significant difference between those who had an ongoing pregnancy at

12 weeks and those who did not have an ongoing pregnancy at 12 weeks regarding the level of FF 25(OH)D and Serum 25(OH)D,  $p$  value  $< 0.001$  (table 11 and figure 4).

#### 3.8.3 Vitamin D for predication of clinical pregnancy

The optimal cutoff point for FF 25(OH)D for prediction of clinical pregnancy was  $\leq 14.7$  ng/ml, the area under the curve was  $0.815$ , the 95% CI was  $0.718-0.890$ ,  $P$  value  $< 0.001$ , Sensitivity  $87.5$ , Specificity  $92.86$ , positive predictive value (PPV)  $87.5$ , negative predictive value (NPV)  $92.9$  and accuracy was  $90.91$ . While the curve analysis for Serum 25(OH)D showed the same figures but with the optimal cutoff point for prediction of clinical pregnancy was  $\leq 23.2$  ng/ml (table 12 and figures 1&2)

## 4. Discussion

The recent advances in Assisted Reproductive Technologies (ART) and bioassays made it possible to study the relationship of both Follicular fluid (FF) and serum levels of vitamin D with the different steps of ICSI including oocyte fertilization, embryo development, and pregnancy outcome. In spite of a number of scientific research which tried to study these relations [25, 31, 32], the relationship between the level of vitamin D (FF and serum) and ICSI outcome(s) remains controversial. It seems that the level of vitamin D has a complex relationship with ICSI outcome. The previous systemic reviews and meta-analysis of serum vitamin D and ICSI outcomes do not support the idea of routinely screening 25(OH)D serum status to predict the clinical pregnancy rate, nor supplementing vitamin D in couples undergoing ART [25, 31, 32]. In this current study, there was a highly statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding the weight and BMI,  $p$  value  $< 0.001$ . These results are in agreement with Rehman R. et al 2013 (34) who reached to the conclusion that a BMI cut off value of above  $26$  kg/m<sup>2</sup> in their study population is associated with a negative impact on pregnancy outcome. However, our results are not in agreement with Adel El Sayed, 2007 [35] who concluded that the overweight and obesity did not affect the ICSI outcome. This discrepancy in the result among different studies could be attributed to the differences in the studied populations. In this study, there was a highly statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding the Day 3 serum FSH,  $p$  value  $< 0.001$ . These findings are in agreement with Shirm, et al 2006, [36], who concluded that elevated day 3 FSH/LH ratio is associated with an inferior outcome in IVF treatment cycles and may be used as an additional predictor for decreased ovarian response. This study also showed that there was a highly statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding Number of retrieved Oocytes, Number of fertilized Oocytes, Number of transferred embryos, Number of frozen Embryos and fertilization rate. These findings are in agreement with Jamal, et al, 2023 [37] and Fanton et al, .2023 [38].

**Table 1:** Demographic data

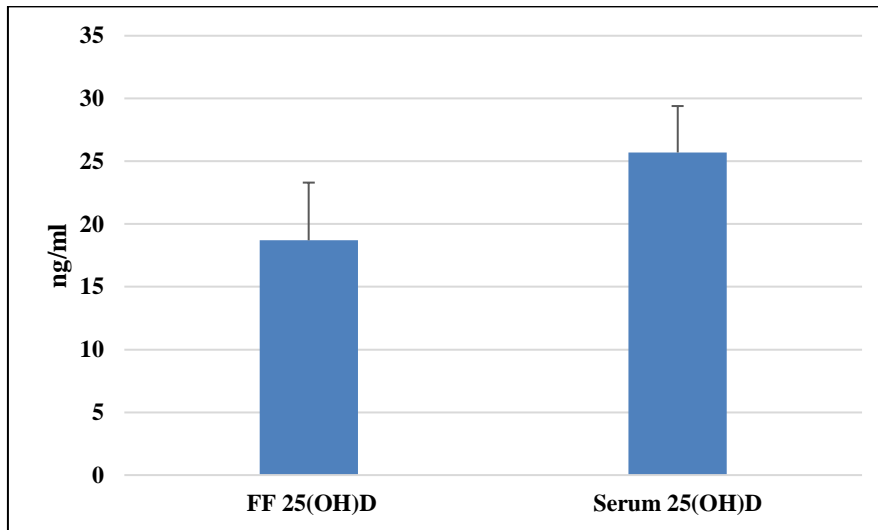
		Descriptive statistics N=88
Age	Range Mean ± SD	(21-38) 28.5±5.3
Weight (KG)	Range Mean ± SD	(45-90.9) 63.5±12.5
Height (M)	Range Mean ± SD	(1.5-1.8) 1.6±0.1
BMI	Range Mean ± SD	(18-29.2) 23.6±3.4
Parity	Median IQR	1 (0-2)
Duration of infertility (Ys)	Median IQR	3 (2-4)

**Table 2:** ICSI data

		Descriptive statistics N=88
Day 3 Serum FSH (IU/L)	Range Mean ± SD	(5.1-11.6) 9.2±2.2
Day of Embryo Transfer	Median IQR	3 (2-3)
	2	40(45.5%)
	3	32(36.4%)
	5	16(18.2%)
Number of Retrieved Oocytes	Median IQR	4 (4-5)
Number of Fertilized Oocytes	Median IQR	3 (2-5)
Fertilization rate	Range Mean ± SD	(50-100) 70.7±14.8
Number of transferred Embryos	Median IQR	1 (1-1)
	1	68(77.3%)
	2	20(22.7%)
Number of Frozen Embryos	Median IQR	1 (1-4)
BHCG on day 14	Median IQR	5 (2-96)
	-Ve	56(63.6%)
	+Ve	32(36.4%)

**Table 3:** Follicular fluid and serum Vitamin D level.

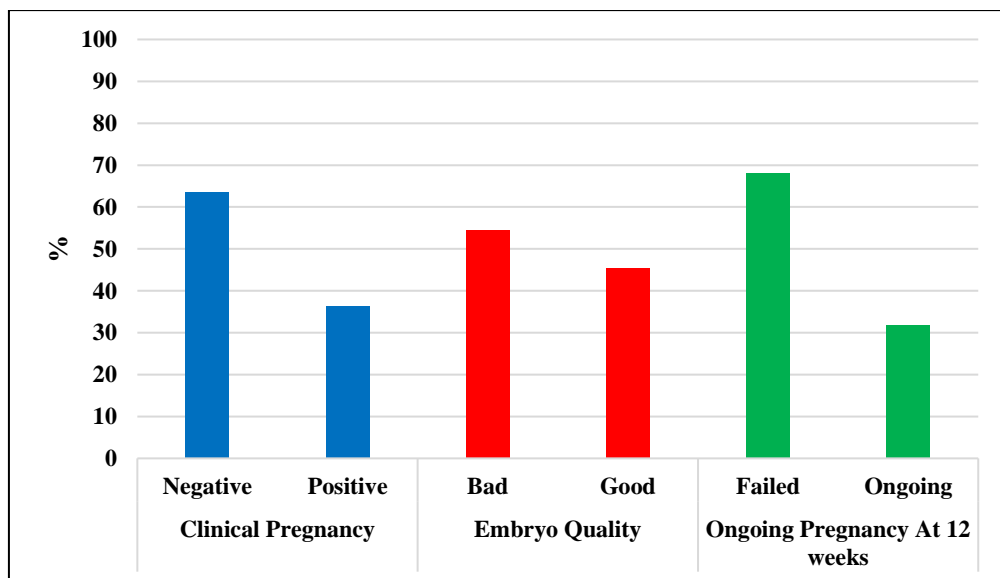
		Descriptive statistics N=88
FF 25(OH)D	Range Mean ± SD	(11.1-25.1) 18.7±4.6
serum 25(OH)D	Range Mean ± SD	(20.1-29.8) 25.7±3.7



**Figure 1:** Follicular fluid and serum Vitamin D level

**Table 4:** Pregnancy outcome

		<b>Descriptive statistics N=88</b>
<b>Clinical Pregnancy</b>	<i>Negative preg. test</i>	56(63.6%)
	<i>Positive preg. test</i>	32(36.4%)
<b>Embryo Quality ((TQE) on the second day of the culture)</b>	<i>Bad quality Emb.</i>	48(54.5%)
	<i>Good Quality Emb.</i>	40(45.5%)
<b>Ongoing Pregnancy At 12 weeks</b>	<i>Failed preg.</i>	60(68.2%)
	<i>Ongoing preg.</i>	28(31.8%)



**Figure 2:** Pregnancy outcome with Clinical pregnancy, Embryo quality and Ongoing pregnancy at 12 weeks

**Table 5:** Correlation between Vitamin D and other clinical and laboratory data

	FF 25(OH)D		Serum 25(OH)D	
	R	P value	r	P value
<b>Serum 25(OH)D<sup>(P)</sup></b>	<b>0.958</b>	<b>&lt;0.001*</b>		
<b>Age<sup>(P)</sup></b>	<b>0.214</b>	<b>0.046*</b>	0.157	0.144
<b>Parity<sup>(P)</sup></b>	<b>-0.284</b>	<b>0.007*</b>	<b>-0.226</b>	<b>0.034*</b>
<b>Duration of infertility (Ys)<sup>(P)</sup></b>	-0.093	0.387	-0.209	0.051
<b>Weight (KG)<sup>(P)</sup></b>	<b>0.373</b>	<b>&lt;0.001*</b>	<b>0.434</b>	<b>&lt;0.001*</b>
<b>Height (M)<sup>(P)</sup></b>	0.182	0.090	<b>0.290</b>	<b>0.006*</b>
<b>BMI<sup>(P)</sup></b>	<b>0.396</b>	<b>&lt;0.001*</b>	<b>0.417</b>	<b>&lt;0.001*</b>
<b>Day 3 Serum FSH (IU/L)<sup>(P)</sup></b>	<b>0.627</b>	<b>&lt;0.001*</b>	<b>0.626</b>	<b>&lt;0.001*</b>
<b>Day of Embryo Transfer<sup>(P)</sup></b>	-0.130	0.228	-0.062	0.568
<b>Number of Retrieved Oocytes<sup>(P)</sup></b>	<b>-0.581</b>	<b>&lt;0.001*</b>	<b>-0.654</b>	<b>&lt;0.001*</b>
<b>Number of Fertilized Oocytes<sup>(P)</sup></b>	<b>-0.592</b>	<b>&lt;0.001*</b>	<b>-0.664</b>	<b>&lt;0.001*</b>
<b>Number of transferred Embryos<sup>(P)</sup></b>	<b>0.345</b>	<b>0.001*</b>	<b>0.315</b>	<b>0.003*</b>
<b>Number of Frozen Embryos<sup>(P)</sup></b>	<b>-0.623</b>	<b>&lt;0.001*</b>	<b>-0.679</b>	<b>&lt;0.001*</b>
<b>BHCG on day 14<sup>(P)</sup></b>	<b>-0.633</b>	<b>&lt;0.001*</b>	<b>-0.656</b>	<b>&lt;0.001*</b>
<b>Fertilization rate<sup>(P)</sup></b>	<b>-0.432</b>	<b>&lt;0.001*</b>	<b>-0.498</b>	<b>&lt;0.001*</b>
<b>Clinical Pregnancy<sup>(S)</sup></b>	<b>-0.525</b>	<b>&lt;0.001*</b>	<b>-0.525</b>	<b>&lt;0.001*</b>
<b>Embryo Quality<sup>(S)</sup></b>	<b>-0.471</b>	<b>&lt;0.001*</b>	<b>-0.450</b>	<b>&lt;0.001*</b>
<b>Ongoing Pregnancy at 12 weeks<sup>(S)</sup></b>	<b>-0.412</b>	<b>&lt;0.001*</b>	<b>-0.435</b>	<b>&lt;0.001*</b>

- (P) Pearson’s correlation
- (S) Spearman’s correlation
- \*: Significant level at P value < 0.05

**Table 6:** Comparison between demographic data and Clinical pregnancy rate

		Clinical Pregnancy		P value
		No	Yes	
		N=56	N=32	
<b>Age</b>	Range Mean ± SD	(21-38) 29.7±5.3	(21-33) 26.2±4.6	<b>0.002*</b>
<b>Weight (KG)</b>	Range Mean ± SD	(45-90.9) 69±11.6	(45-64.6) 53.8±6.8	<b>&lt;0.001*</b>
<b>Height (M)</b>	Range Mean ± SD	(1.6-1.8) 1.6±0.1	(1.5-1.7) 1.6±0.1	<b>0.007*</b>
<b>BMI</b>	Range Mean ± SD	(18-29.2) 25.3±3.1	(18.7-22.9) 20.7±1.2	<b>&lt;0.001*</b>
<b>Parity</b>	Median IQR	1 (0-2)	1 (0.3-2)	0.459
<b>Duration of infertility (Ys)</b>	Median IQR	3 (2-4)	3 (3-4)	0.427

- Independent Samples T test for parametric quantitative data between the two groups
- Mann Whitney test for non-parametric quantitative data between the two groups
- \*: Significant level at P value < 0.05

**Table 7:** Comparison between ICSI data and Clinical pregnancy rate.

		Clinical Pregnancy		P value
		No	Yes	
		N=56	N=32	
<b>Day 3 Serum FSH (IU/L)</b>	<i>Range</i> <i>Mean ± SD</i>	(9.1-11.6) 10.8±0.6	(5.1-7.5) 6.4±0.7	<0.001*
<b>Day of Embryo Transfer</b>	<i>Median</i> <i>IQR</i>	2 (2-3)	3 (2.3-4.5)	0.007*
	2	32(57.1%)	8(25%)	0.014*
	3	16(28.6%)	16(50%)	
	5	8(14.3%)	8(25%)	
<b>Number of Retrieved Oocytes</b>	<i>Median</i> <i>IQR</i>	4 (3-4)	6 (5-6.8)	<0.001*
<b>Number of Fertilized Oocytes</b>	<i>Median</i> <i>IQR</i>	2 (2-3)	5 (4.3-5.8)	<0.001*
<b>Number of transferred Embryos</b>	<i>Median</i> <i>IQR</i>	1 (1-2)	1 (1-1)	<0.001*
	1	36(64.3%)	32(100%)	<0.001*
	2	20(35.7%)	0(0%)	
<b>Number of Frozen Embryos</b>	<i>Median</i> <i>IQR</i>	1 (1-1)	4 (3.3-4.8)	<0.001*
<b>Fertilization rate</b>	<i>Range</i> <i>Mean ± SD</i>	(50-75) 62.6±10.4	(71.4-100) 84.9±9.9	<0.001*

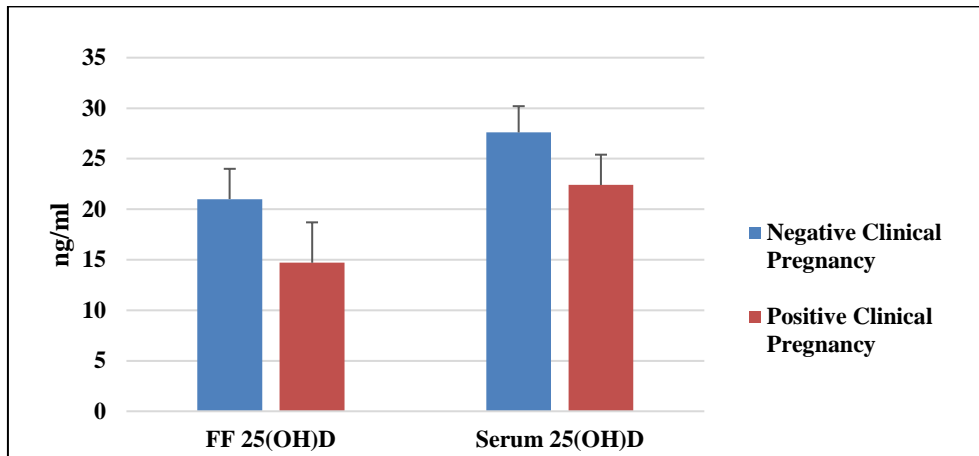
- Independent Samples T test for parametric quantitative data between the two groups
- Mann Whitney test for non-parametric quantitative data between the two groups
- \*: Significant level at P value < 0.05

**Table 8:** Comparison between Vitamin D and Clinical pregnancy rate.

		Clinical Pregnancy		P value
		No	Yes	
		N=56	N=32	
<b>FF 25(OH)D</b>	<i>Range</i> <i>Mean ± SD</i>	(11.1-25.1) 21±3	(11.4-25.1) 14.7±4	<0.001*
<b>Serum 25(OH)D</b>	<i>Range</i> <i>Mean ± SD</i>	(20.1-29.8) 27.6±2.6	(20.3-29.8) 22.4±3	<0.001*

- Independent Samples T test for parametric quantitative data between the two groups
- \*: Significant level at P value < 0.05



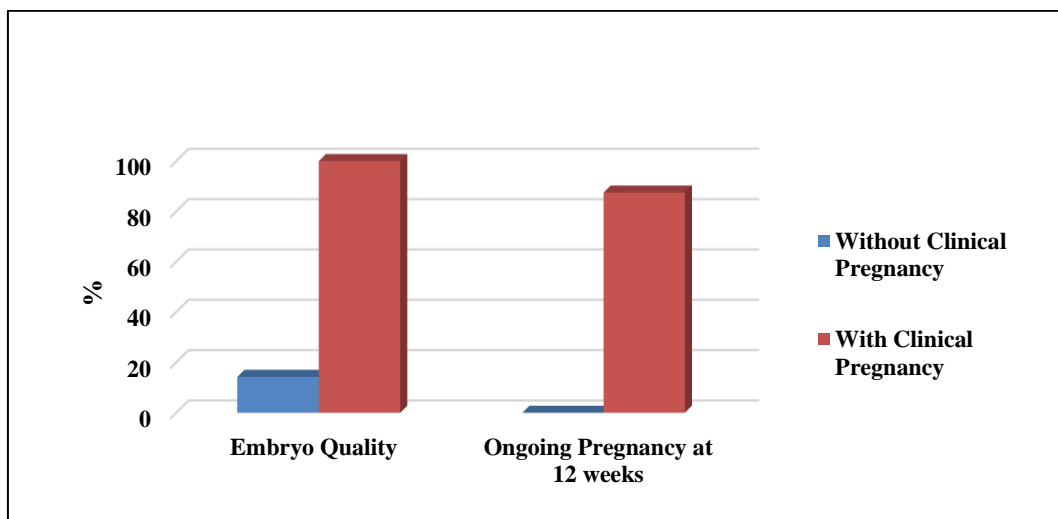


**Figure 3:** Vitamin D and Clinical pregnancy

**Table 9:** Comparison between Embryo quality, ongoing pregnancy, and Clinical pregnancy rate.

		Clinical Pregnancy		P value
		No	Yes	
		N=56	N=32	
<b>Embryo Quality</b>	No	48(85.7%)	0(0%)	<b>&lt;0.001*</b>
	Yes	8(14.3%)	32(100%)	
<b>Ongoing Pregnancy at 12 weeks</b>	No	56(100%)	4(12.5%)	<b>&lt;0.001*</b>
	Yes	0(0%)	28(87.5%)	

- Chi square test for qualitative data between the two groups
- \*: Significant level at P value < 0.05

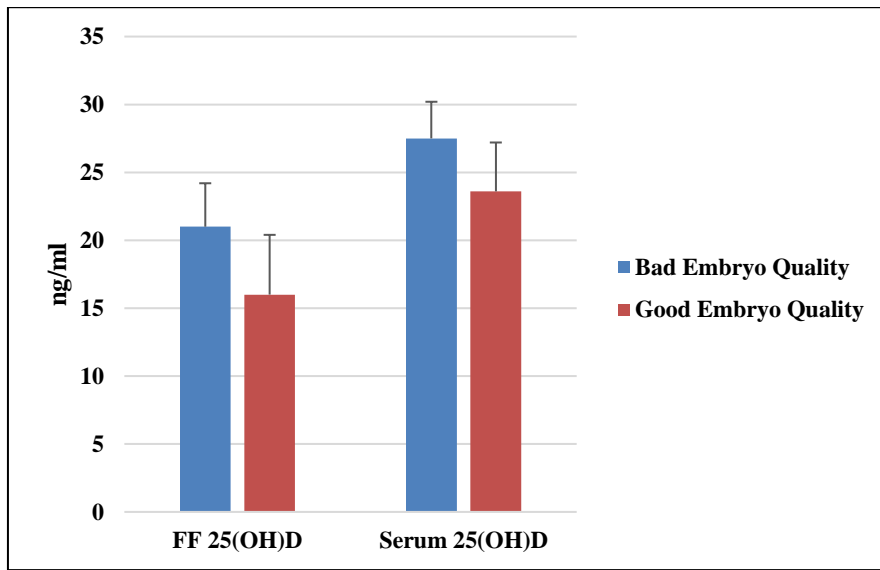


**Figure 4:** Embryo quality, ongoing pregnancy, and Clinical pregnancy

**Table 10:** Comparison between Vitamin D and embryo quality.

		Embryo Quality		P value
		No	Yes	
		N=48	N=40	
<b>FF 25(OH)D</b>	Range Mean ± SD	(11.1-25.1) 21±3.2	(11.4-25.1) 16±4.4	<0.001*
<b>Serum 25(OH)D</b>	Range Mean ± SD	(20.1-29.8) 27.5±2.7	(20.3-29.8) 23.6±3.6	<0.001*

- Independent Samples T test for parametric quantitative data between the two groups
- \*: Significant level at P value < 0.05

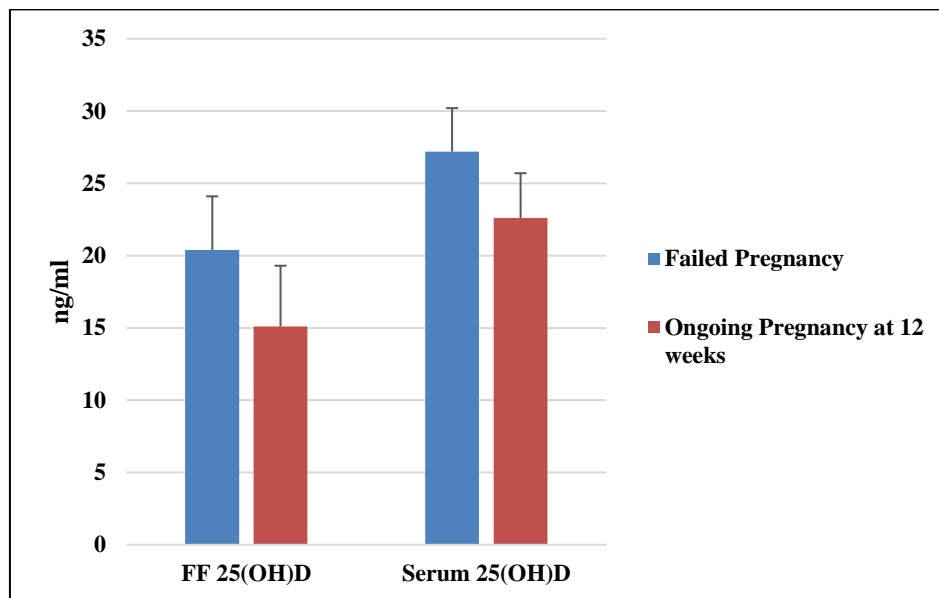


**Figure 5:** Follicular Fluid and serum Vitamin D and embryo quality

**Table 11:** Comparison between Vitamin D and ongoing pregnancy at 12 weeks.

		Ongoing Pregnancy at 12 weeks		P value
		No	Yes	
		N=60	N=28	
<b>FF 25(OH)D</b>	Range Mean ± SD	(11.1-25.1) 20.4±3.7	(11.4-25.1) 15.1±4.2	<0.001*
<b>Serum 25(OH)D</b>	Range Mean ± SD	(20.1-29.8) 27.2±3	(20.3-29.8) 22.6±3.1	<0.001*

- Independent Samples T test for parametric quantitative data between the two groups
- \*: Significant level at P value < 0.05



**Figure 6:** Comparison between Vitamin D and ongoing pregnancy at 12 weeks

Although these two groups of investigators were working on IVF cycles, however, these findings could be extended to ICSI cycles as well. This study also showed that there was a statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding the Day of embryo transfer (day five transfer was the best,  $p$  value = 0.014). These findings are in agreement with Tökmeci et al., 2017 [39]. In this current study, FF 25(OH) D was lower than that of Serum 25(OH)D. These findings are in agreement with Jeremic et al., 2021 (40) and Ozyurt R & Karaku C., 2022 [12], but not in agreement with Ciepiela, et al., 2018, [41]. In this study, there was a highly statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding the FF 25(OH) D and Serum 25(OH) D,  $P$  value < 0.001. These findings are in agreement with Garbedian, et al. 2013 [42], who suggested that women with sufficient levels of vitamin D are significantly more likely to achieve clinical pregnancy following IVF. Garbedian et al, 2013, differ from this current study in that they only measured serum 25(OH) D and that they adopted IVF not ICSI protocol. In this current study, there was a highly statistically significant difference between those who got clinical pregnancy and those who did not get clinical pregnancy regarding the Embryo quality and ongoing pregnancy at 12 weeks,  $P$  value < 0.001. These findings are not in agreement with Oron et al., 2014 [43]. However, the later study was a retrospective study. In this current study, There was a highly statistically significant difference between those who had a good Embryo quality and those who did not have good Embryo quality regarding the level of FF 25(OH) D and Serum 25(OH) D,  $p$  value < 0.001. These findings are in a partial agreement with those of Abadia et al, 2016 [44]. In this study, There was a highly statistically significant difference between those who had an ongoing pregnancy at 12 weeks and those who did not have an ongoing pregnancy at 12 weeks regarding the level of FF

25(OH) D and Serum 25(OH) D ,  $p$  value < 0.001. These findings are not in agreement with Álvarez-Silvares et al , 2016 (45) who concluded that The pregnancy outcome was independent of the first trimester maternal serum 25(OH)D status. This disagreement could be attributed to the differences in studied population. To focus on the evaluation of oocyte competence, we excluded couples with moderate and severe male factor and therefore minimized the role of spermatozoa as a factor affecting embryo development. In contrast, in some previous studies, male factor was present in 58–65% of couples (14, 24), while in other studies (16, 19, and 24), the indications for IVF/ ICSI were not mentioned. Additionally, to test only mature oocytes, we evaluated only patients undergoing ICSI.

## 5. Conclusions

Follicular Fluid 25(OH) D and Serum 25(OH) D levels are markers of oocyte quality and a marker of clinical pregnancy rate after ICSI.

## References

- [1] M. Holick. (2007). Vitamin D deficiency New England Journal of Medicine. 357: 266–81 doi: 10.1056. NEJMra070553.
- [2] J. Luk, S. Torrealday, G. Neal Perry, L. Pal. (2012). Relevance of vitamin D in reproduction. Human reproduction. 27(10): 3015-3027.
- [3] J.A. Johnson, J.P. Grande, P.C. Roche, R. Kumar. (1996). Immunohistochemical detection and distribution of the 1, 25-dihydroxyvitamin D 3 receptor in rat reproductive tissues. Histochemistry and cell biology. 105: 7-15.
- [4] J. Wojtusik, P.A. Johnson. (2012). Vitamin D regulates anti-Mullerian hormone expression in granulosa cells of the hen. Biology of reproduction. 86(3): 91, 1-7.
- [5] L.M. Bodnar, J.M. Catov, H.N. Simhan, M.F. Holick, R.W. Powers, J.M. Roberts. (2007).

- Maternal vitamin D deficiency increases the risk of preeclampsia. *The Journal of Clinical Endocrinology & Metabolism*. 92(9): 3517-3522.
- [6] F. Aghajafari, T. Nagulesapillai, P.E. Ronksley, S.C. Tough, M. O'Beirne, D.M. Rabi. (2013). Association between maternal serum 25-hydroxyvitamin D level and pregnancy and neonatal outcomes: systematic review and meta-analysis of observational studies. *British medical journal*. 346.
- [7] E. Wehr, T. Pieber, B. Obermayer-Pietsch. (2011). Effect of vitamin D3 treatment on glucose metabolism and menstrual frequency in polycystic ovary syndrome women: a pilot study. *Journal of endocrinological investigation*. 34: 757-763.
- [8] M. Asadi, N. Matin, M. Frootan, J. Mohamadpour, M. Qorbani, F.D. Tanha. (2014). Vitamin D improves endometrial thickness in PCOS women who need intrauterine insemination: a randomized double-blind placebo-controlled trial. *Archives of gynecology and obstetrics*. 289: 865-870.
- [9] J. Ott, L. Wattar, C. Kurz, R. Seemann, J. Huber, K. Mayerhofer, E. Vytiska-Binstorfer. (2012). Parameters for calcium metabolism in women with polycystic ovary syndrome who undergo clomiphene citrate stimulation: a prospective cohort study. *European journal of endocrinology*. 166(5): 897-902.
- [10] A. Lasco, A. Catalano, S. Benvenga. (2012). Improvement of primary dysmenorrhea caused by a single oral dose of vitamin D: results of a randomized, double-blind, placebo-controlled study. *Archives of internal medicine*. 172(4): 366-367.
- [11] L.A. Wise, E.A. Ruiz-Narváez, S.A. Haddad, L. Rosenberg, J.R. Palmer. (2014). Polymorphisms in vitamin D-related genes and risk of uterine leiomyomata. *Fertility and sterility*. 102(2): 503-510. e1.
- [12] R. Ozyurt, C. Karakus. (2022). Follicular fluid 25-hydroxyvitamin D levels determine fertility outcome in patients with polycystic ovary syndrome. *Taiwanese Journal of Obstetrics and Gynecology*. 61(4): 620-625.
- [13] N.P. Polyzos, E. Anckaert, L. Guzman, J. Schiettecatte, L. Van Landuyt, M. Camus, J. Smits, H. Tournaye. (2014). Vitamin D deficiency and pregnancy rates in women undergoing single embryo, blastocyst stage, transfer (SET) for IVF/ICSI. *Human reproduction*. 29(9): 2032-2040.
- [14] R.D. Firouzabadi, E. Rahmani, M. Rahsepar, M.M. Firouzabadi. (2014). Value of follicular fluid vitamin D in predicting the pregnancy rate in an IVF program. *Archives of gynecology and obstetrics*. 289: 201-206.
- [15] J.M. Franasiak, T.A. Molinaro, E.K. Dubell, K.L. Scott, A.R. Ruiz, E.J. Forman, M.D. Werner, K.H. Hong, R.T. Scott Jr. (2015). Vitamin D levels do not affect IVF outcomes following the transfer of euploid blastocysts. *American journal of obstetrics and gynecology*. 212(3): 315. e1-315. e6.
- [16] S. Ozkan, S. Jindal, K. Greenseid, J. Shu, G. Zeitlian, C. Hickmon, L. Pal. (2010). Replete vitamin D stores predict reproductive success following in vitro fertilization. *Fertility and sterility*. 94(4): 1314-1319.
- [17] B. Rudick, S. Ingles, K. Chung, F. Stanczyk, R. Paulson, K. Bendikson. (2012). Characterizing the influence of vitamin D levels on IVF outcomes. *Human reproduction*. 27(11): 3321-3327.
- [18] A. Paffoni, S. Ferrari, P. Viganò, L. Pagliardini, E. Papaleo, M. Candiani, A. Tirelli, L. Fedele, E. Somigliana. (2014). Vitamin D deficiency and infertility: insights from in vitro fertilization cycles. *The Journal of Clinical Endocrinology & Metabolism*. 99(11): E2372-E2376.
- [19] A. Aleyasin, M.A. Hosseini, A. Mahdavi, L. Safdarian, P. Fallahi, M.R. Mohajeri, M. Abbasi, F. Esfahani. (2011). Predictive value of the level of vitamin D in follicular fluid on the outcome of assisted reproductive technology. *European Journal of Obstetrics & Gynecology and Reproductive Biology*. 159(1): 132-137.
- [20] A.M. Fabris, M. Cruz, C. Iglesias, A. Pacheco, A. Patel, J. Patel, H. Fatemi, J.A. García-Velasco. (2017). Impact of vitamin D levels on ovarian reserve and ovarian response to ovarian stimulation in oocyte donors. *Reproductive biomedicine online*. 35(2): 139-144.
- [21] C. Bagot, P. Troy, H. Taylor. (2000). Alteration of maternal Hoxa10 expression by in vivo gene transfection affects implantation. *Gene Therapy*. 7(16): 1378-1384.
- [22] H. Du, G.S. Daftary, S.I. Lalwani, H.S. Taylor. (2005). Direct regulation of HOXA10 by 1, 25-(OH) 2D3 in human myelomonocytic cells and human endometrial stromal cells. *Molecular endocrinology*. 19(9): 2222-2233.
- [23] G.M. Anifandis, K. Dafopoulos, C.I. Messini, N. Chalvatzas, N. Liakos, S. Pournaras, I.E. Messinis. (2010). Prognostic value of follicular fluid 25-OH vitamin D and glucose levels in the IVF outcome. *Reproductive Biology and Endocrinology*. 8(1): 1-5.
- [24] L. Farzadi, H.K. Bidgoli, M. Ghojzadeh, Z. Bahrami, A. Fattahi, Z. Latifi, V. Shahnazi, M. Nouri. (2015). Correlation between follicular fluid 25-OH vitamin D and assisted reproductive outcomes. *Iranian journal of reproductive medicine*. 13(6): 361.
- [25] L.E. Johnson, H.F. DeLuca. (2001). Vitamin D receptor null mutant mice fed high levels of calcium are fertile. *The Journal of nutrition*. 131(6): 1787-1791.
- [26] M.M. Pacis, C.N. Fortin, S.M. Zarek, S.L. Mumford, J.H. Segars. (2015). Vitamin D and assisted reproduction: should vitamin D be routinely screened and repleted prior to ART? A systematic review. *Journal of assisted reproduction and genetics*. 32: 323-335.
- [27] D.A. Holick, R.P. Heaney, et al. (2011). Evaluation, treatment, and prevention of vitamin D deficiency: an endocrine society clinical practice guideline. *The Journal of clinical endocrinology and metabolism*. 96:1911-30.

- [28] C.J. Rosen, S.A. Abrams, J.F. Aloia, P.M. Brannon, S.K. Clinton, R.A. Durazo-Arvizu, J.C. Gallagher, R.L. Gallo, G. Jones, C.S. Kovacs. (2012). IOM committee members respond to Endocrine Society vitamin D guideline. *The Journal of Clinical Endocrinology & Metabolism*. 97(4): 1146-1152.
- [29] P. Ciepiela, T. Bączkowski, A. Drozd, A. Kazienko, E. Stachowska, R. Kurzawa. (2015). Arachidonic and linoleic acid derivatives impact oocyte ICSI fertilization—a prospective analysis of follicular fluid and a matched oocyte in a ‘one follicle—one retrieved oocyte—one resulting embryo’investigational setting. *PLoS One*. 10(3): e0119087.
- [30] M.P. Rosen, S. Shen, A.T. Dobson, V.Y. Fujimoto, C.E. McCulloch, M.I. Cedars. (2006). Oocyte degeneration after intracytoplasmic sperm injection: a multivariate analysis to assess its importance as a laboratory or clinical marker. *Fertility and sterility*. 85(6): 1736-1743.
- [31] B. Balaban, D. Brison, G. Calderon, J. Catt, J. Conaghan, L. Cowan, T. Ebner, D. Gardner, T. Hardarson, K. Lundin. (2011). Istanbul consensus workshop on embryo assessment: proceedings of an expert meeting. *Reproductive biomedicine online*. 22(6): 632-646.
- [32] A.C. Miklos, C. Li, C.D. Sorrell, L.A. Lyon, G.J. Pielak. (2011). An upper limit for macromolecular crowding effects. *BMC biophysics*. 4: 1-7.
- [33] S.S. Lv, J.Y. Wang, X.Q. Wang, Y. Wang, Y. Xu. (2016). Serum vitamin D status and in vitro fertilization outcomes: a systematic review and meta-analysis. *Archives of gynecology and obstetrics*. 293: 1339-1345.
- [34] R. Rehman, Z. Hussain, S.S. Fatima. (2013). Effect of weight status on pregnancy outcome in intra cytoplasmic sperm injection. *Iranian journal of reproductive medicine*. 11(9): 717.
- [35] A. El Sayed. (2007). THE MINIMAL EFFECT OF BODY MASS INDEX ON ICSI OUTCOME. *The Egyptian Journal of Fertility of Sterility*. 11(2): 35-40.
- [36] A. Shrim, S. Elizur, D. Seidman, J. Rabinovici, A. Wisner, J. Dor. (2006). Elevated day 3 FSH/LH ratio due to low LH concentrations predicts reduced ovarian response. *Reproductive biomedicine online*. 12(4): 418-422.
- [37] M. Jamil, H. Debbarh, A. Kabit, M. Ennaji, M. Zargaoui, W. Senhaji, M. Hissane, B. Saadani, N. Louanjli, R. Cadi. (2023). Impact of the number of retrieved oocytes on IVF outcomes: oocyte maturation, fertilization, embryo quality and implantation rate. *Zygote*. 31(1): 91-96.
- [38] M. Fanton, J.H. Cho, V.L. Baker, K. Loewke. (2023). A higher number of oocytes retrieved is associated with an increase in fertilized oocytes, blastocysts, and cumulative live birth rates. *Fertility and sterility*. 119(5): 762-769.
- [39] Tökmeci, Ö , Dogan, Muammer , Topçu, Hasan Onur , Demirel, Ö . Guzel, Ali. (2017). Effect of day 3 or 5 embryo transfer on pregnancy rates and perinatal outcomes: Evaluation of 1,291 ICSI/IVF Cycles. *Journal of Reproductive Medicine*. 62. 653-658.
- [40] A. Jeremic, Z. Mikovic, I. Soldatovic, E. Sudar-Milovanovic, E.R. Isenovic, M. Perovic. (2021). Follicular and serum levels of vitamin D in women with unexplained infertility and their relationship with in vitro fertilization outcome: an observational pilot study. *Archives of Medical Science: AMS*. 17(5): 1418.
- [41] P. Ciepiela, A.J. Dulęba, E. Kowaleczko, K. Chelstowski, R. Kurzawa. (2018). Vitamin D as a follicular marker of human oocyte quality and a serum marker of in vitro fertilization outcome. *Journal of assisted reproduction and genetics*. 35: 1265-1276.
- [42] K. Garbedian, M. Boggild, J. Moody, K.E. Liu. (2013). Effect of vitamin D status on clinical pregnancy rates following in vitro fertilization. *Canadian Medical Association Open Access Journal*. 1(2): E77-E82.
- [43] G. Oron, W.-Y. Son, W. Buckett, T. Tulandi, H. Holzer. (2014). The association between embryo quality and perinatal outcome of singletons born after single embryo transfers: a pilot study. *Human reproduction*. 29(7): 1444-1451.
- [44] L. Abadia, Gaskins A ,J , Chiu Y Williams P L, Myra Keller M , Wright D L ,Souter I, Hauser R, Chavarro J E. (2016). Serum 25-hydroxyvitamin D concentrations and treatment outcomes of women undergoing assisted reproduction. *The American Journal of Clinical Nutrition*. 104:729–35.
- [45] E. Álvarez-Silvares, M. Vilouta-Romero, E. Borrajo-Hernández, M.L. Morales-Serrano, M.T. Alves-Pérez. (2017). Concentraciones séricas maternas de 25-hidroxivitamina D en el primer trimestre y resultados adversos gestacionales. *Ginecología y Obstetricia de México*. 84(03): 150-163.